

# Preservation of Fungi in an Atmosphere over Liquid Nitrogen after Uncontrolled Freezing

Toshirou Nagai\*, Aika Ideno, Michiyo Tsuge, Chie Oyanagi\*\*, Masaomi Oniki, Kouichi Kita\*\*\*, Mitsuo Horita, Takayuki Aoki, Takao Kobayashi and Kenichi Tsuchiya

National Institute of Agrobiological Resources, Ministry of Agriculture, Forestry and Fisheries, Tsukuba, Ibaraki 305-8602, Japan

In the Genebank of the Ministry of Agriculture, Forestry and Fisheries in Japan, 5,406 (95.7%) of 5,651 maintained fungal strains (466 genera and 1,182 species), including strains of oomycetes, were well stored in an atmosphere over liquid nitrogen at  $-165^{\circ}\text{C}$  after uncontrolled freezing, showing that this method is applicable to preservation of most of fungal strains. Percentages of strains stored successfully for 1 year within Oomycota, Zygomycota, Ascomycota, Basidiomycota, and anamorphic fungi, were 59.3% (233/393 strains tested), 98.3% (58/59), 98.8% (881/892), 95.4% (1,100/1,153), and 99.4% (3,134/3,154), respectively. Strains of *Achlya*, *Dictyuchus* (Oomycota), *Volvariella* (Basidiomycota), and *Pseudoseptoria* (anamorphic fungi) could not be stored using this method. Successful preservation depends on the species and strains of fungi *sensu lato*.

Key words : cryopreservation, fungi

In many culture collections, fungal strains are maintained using several methods, such as freeze-drying, oil storage, and cryopreservation (7, 8). The Genebank of the Ministry of Agriculture, Forestry and Fisheries (MAFF) in Japan has collected about 9,000 fungal isolates (including oomycetes) which are stored mainly in an atmosphere over liquid nitrogen after uncontrolled freezing, because this provides stable preservation without the need for complicated handling. Frozen strains have been tested for their viability at intervals, and data concerning viability of stored strains have been accumulated. In this report, viability after one-year

storage of the preserved strains belonging to Oomycota, Zygomycota, Ascomycota, Basidiomycota, and anamorphic fungi, is summarized. The subdivisions are used according to Hawksworth et al. (1); however, "fungi" include oomycetes in this report.

Fungal strains tested are listed in alphabetical order of generic names (Table 1). The culture medium used for preparing stocks and testing viability was potato dextrose agar (Difco), for most of the species. V8 juice agar (9) was used for oomycetes. V8 juice agar contained 20% of the supernatant of commercial V8 juice (Campbell Soup Company, Camden, NJ), which was obtained by centrifugation of a mixture of the juice and 1.5%  $\text{CaCO}_3$  (9).

Preservation of fungi was performed according to the method of Miki and Kubomura (3), with some modifications. Fungal strains were grown on the agar media for 1-2 weeks at  $20^{\circ}\text{C}$ . Agar disks (6mm dia.,  $70\mu\text{l}$ ) with spores and/or mycelia were cut off from a plate using gas-sterilized plastic straws. Five or ten disks were soaked in 1ml of 10% glycerol in a 2-ml plastic vial. The vial was allowed to stand at

\* Corresponding author : E-mail ; nag@affrc.go.jp

\*\* Present address : National Center for Seeds and Seedlings, Agriculture, Forestry and Fisheries Research Council, Ministry of Agriculture, Forestry and Fisheries, Tsukuba, Ibaraki 305-0852, Japan

\*\*\* Present address : Hokkaido National Agricultural Experiment Station, Ministry of Agriculture, Forestry and Fisheries, Sapporo, Hokkaido 062-8555, Japan

5°C for 2 days, for cold-hardening, and then at -60°C in an electrical deep freezer for 2 days. Finally, the vial was transferred to an atmosphere over liquid nitrogen at -165°C.

After 1 month (0-2 months) and 1 year (9.6-14.4 months), a vial containing frozen fungus was thawed quickly in a water bath, either at 30°C (5) or at 37°C (3), for 5 min. All disks were placed onto suitable agar for each strain and incubated at the appropriate growing temperature. Viability was defined as the percentage of disks on which visible growth was observed, and viability of more than 80% was assessed as successful preservation. A computer software; seizan. exe (4), was used to process viability data.

Among 5,651 fungal strains of 466 genera and 1,182 species, 5,503 strains (97.4% of total strains) were recovered successfully after storage for 1 month and 5,406 strains (95.7%) after 1 year (Table 1). Percentages of strains stored successfully for 1 year within Oomycota, Zygomycota, Ascomycota, Basidiomycota, and anamorphic fungi, were 59.3% (233/393 tested strains), 98.3% (58/59), 98.8% (881/892), 95.4% (1,100/1,153), and 99.4% (3,134/3,154), respectively. While most of the fungal strains could be stored by the uncontrolled freezing method, some strains could not be preserved by this method. No strains of the genera *Achlya* (Oomycota, 15 strains tested), *Dictyuchus* (Oomycota, 6 strains), *Volvariella* (Basidiomycota, 1 strain), and *Pseudoseptoria* (anamorphic fungi, 1 strain) survived one-year storage (Table 1). These strains could be stored by controlled freezing (5). Suitable conditions for preservation of strains that are not stored successfully by uncontrolled freezing are under investigation, and therefore these strains are currently maintained by subculturing on agar slants occasionally with oil overlay.

Though species of *Phytophthora* and *Pythium* are known to have low survival after long term storage (6), some strains of the genera were found alive. Successful preservation depended on species or strains. Some strains of *Phytophthora irregulare* (24 surviving strains/28 strains tested) and *Pythium sylvaticum* (8/10) survived well, whereas all the examined strains of *Py. oedochilum* and *Py. vexans* did not survive one-year storage. The detailed results of the species stored unsuccessfully are given in Table 2.

**Table 1. Number of fungal strains surviving by the cryopreservation**

Fungal genus	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>
Oomycota			
<i>Achlya</i>	15	1	0
<i>Aphanomyces</i>	6	2	2
<i>Dictyuchus</i>	6	0	0
<i>Phytophthora</i>	104	88	78
<i>Plectospora</i>	1	1	1
<i>Pythium</i>	255	177	149
<i>Saprolegnia</i>	2	2	2
<i>Thraustotheca</i>	4	1	1
Zygomycota			
<i>Absidia</i>	3	3	3
<i>Choanephora</i>	2	1	1
<i>Cunninghamella</i>	2	2	2
<i>Gongronella</i>	2	2	2
<i>Mortierella</i>	33	33	33
<i>Mucor</i>	8	8	8
<i>Rhizopus</i>	7	7	7
<i>Syncephalastrum</i>	2	2	2
Ascomycota			
<i>Acrospermum</i>	12	12	12
<i>Amphiportha</i>	1	1	1
<i>Amphisphaeria</i>	1	1	1
<i>Apiospora</i>	3	3	3
<i>Arenariomyces</i>	17	17	17
<i>Ascocalyx</i>	5	5	5
<i>Botryosphaeria</i>	30	30	30
<i>Bulgaria</i>	1	1	1
<i>Calonectria</i>	13	13	13
<i>Calycellina</i>	1	1	1
<i>Ceratocystis</i>	2	2	2
<i>Ceriosporopsis</i>	39	39	39
<i>Chaetomium</i>	22	22	22
<i>Ciborinia</i>	2	2	2
<i>Cistella</i>	19	19	16
<i>Claviceps</i>	7	7	7
<i>Corollospora</i>	33	33	33
<i>Cryphonectria</i>	46	46	46
<i>Cryptodiaportha</i>	3	3	3
<i>Cucurbitodithis</i>	1	1	1
<i>Daldinia</i>	1	1	1
<i>Diaportha</i>	40	40	40
<i>Diaporthopsis</i>	1	1	1
<i>Diatrype</i>	4	4	4
<i>Diatrypella</i>	1	1	1
<i>Didymella</i>	14	14	14

Table 1. continued.

Fungal genus	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>
<i>Discostroma</i>	1	1	1
<i>Dothidea</i>	2	2	2
<i>Endothia</i>	1	1	1
<i>Epichloe</i>	10	10	10
<i>Eudarlucia</i>	2	2	2
<i>Eurotium</i>	2	2	2
<i>Gaeumannomyces</i>	10	10	10
<i>Gibberella</i>	2	2	2
<i>Glomerella</i>	33	33	33
<i>Gnomonia</i>	1	1	1
<i>Graphostroma</i>	6	6	6
<i>Gremmeniella</i>	9	9	8
<i>Grovesinia</i>	4	4	4
<i>Guignardia</i>	38	38	38
<i>Gyromitra</i>	1	1	1
<i>Halbania</i>	1	1	1
<i>Halosarpheia</i>	15	15	15
<i>Halosphaeria</i>	18	18	17
<i>Halosphaeriopsis</i>	30	30	30
<i>Hypomyces</i>	11	11	11
<i>Hypoxylon</i>	13	13	13
<i>Khuskia</i>	1	1	1
<i>Lachnellula</i>	7	7	7
<i>Leptosphaeria</i>	1	1	1
<i>Leptosphaerulina</i>	6	6	6
<i>Leucostoma</i>	2	2	2
<i>Lophiosphaerella</i>	1	1	1
<i>Lophodermium</i>	2	2	2
<i>Marinospora</i>	9	9	9
<i>Massarinula</i>	1	1	1
<i>Melanconis</i>	8	8	8
<i>Microascus</i>	1	1	1
<i>Mixia</i>	1	1	1
<i>Monascus</i>	14	14	14
<i>Monilinia</i>	10	10	10
<i>Monosporascus</i>	2	2	2
<i>Morchella</i>	3	3	2
<i>Mycosphaerella</i>	28	28	28
<i>Nectria</i>	13	13	13
<i>Neocosmospora</i>	2	2	2
<i>Ophiostoma</i>	15	15	15
<i>Ophiovalsa</i>	3	3	3
<i>Pezicula</i>	4	4	4
<i>Peziza</i>	4	3	1
<i>Phaeosphaerella</i>	1	1	1
<i>Phyllachora</i>	1	1	1
<i>Plagiosphaera</i>	1	1	1

Table 1. continued.

Fungal genus	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>
<i>Plectosphaera</i>	6	6	6
<i>Pleospora</i>	4	4	4
<i>Pseudovalsa</i>	5	5	5
<i>Pseudovalsella</i>	1	1	1
<i>Pyrenophora</i>	2	2	2
<i>Remispora</i>	25	25	24
<i>Rhizina</i>	4	4	4
<i>Rosellinia</i>	25	25	25
<i>Rosenscheldiella</i>	1	1	1
<i>Sarea</i>	20	20	20
<i>Sclerotinia</i>	45	45	44
<i>Septotinia</i>	4	4	4
<i>Sordaria</i>	1	1	1
<i>Sphaerulina</i>	1	1	1
<i>Talaromyces</i>	1	1	1
<i>Taphrina</i>	9	9	9
<i>Thielavia</i>	2	2	2
<i>Torpedospora</i>	20	20	20
<i>Trichocoma</i>	1	1	1
<i>Trichoscyphella</i>	4	4	4
<i>Valsa</i>	32	32	32
<i>Valsella</i>	1	1	1
<i>Venturia</i>	7	7	7
<i>Xylaria</i>	1	1	1
Basidiomycota			
<i>Abortiporus</i>	2	2	2
<i>Agaricus</i>	6	6	6
<i>Agrocybe</i>	3	3	3
<i>Amanita</i>	2	2	1
<i>Amylostereum</i>	2	2	2
<i>Anthracoephyllum</i>	3	3	3
<i>Antrodia</i>	11	11	11
<i>Antrodiella</i>	3	3	3
<i>Armillaria</i>	56	54	54
<i>Asterophora</i>	3	3	3
<i>Auricularia</i>	16	14	12
<i>Bjerkandera</i>	5	5	5
<i>Bondarzewia</i>	1	1	1
<i>Botryobasidium</i>	1	1	1
<i>Calocera</i>	4	4	4
<i>Calvatia</i>	4	4	4
<i>Ceratobasidium</i>	49	46	37
<i>Cerrena</i>	1	1	1
<i>Chlorophyllum</i>	1	1	1
<i>Clavicornia</i>	1	1	1
<i>Clitocybe</i>	2	2	2
<i>Collybia</i>	6	6	6

Table 1. continued.

Fungal genus	Tested <sup>(a)</sup>	1 M <sup>(b)</sup>	1 Y <sup>(b)</sup>
<i>Coniophora</i>	1	1	1
<i>Coprinus</i>	10	10	9
<i>Crinipellis</i>	1	1	1
<i>Cryptoporus</i>	3	3	3
<i>Cylindrobasidium</i>	1	1	1
<i>Cyptotrama</i>	2	2	2
<i>Dacrymyces</i>	5	5	5
<i>Dacryopinax</i>	3	3	3
<i>Daedalea</i>	4	4	4
<i>Daedaleopsis</i>	3	3	3
<i>Datronia</i>	2	2	2
<i>Dictyopanus</i>	6	6	6
<i>Diplomitoporus</i>	2	2	2
<i>Ditiola</i>	1	1	1
<i>Eichleriella</i>	10	10	8
<i>Erythricium</i>	18	17	17
<i>Exidia</i>	1	1	1
<i>Exidiopsis</i>	1	1	1
<i>Exobasidium</i>	22	22	22
<i>Favolaschia</i>	1	1	1
<i>Femsjonia</i>	3	3	3
<i>Filoboletus</i>	1	1	1
<i>Fistulina</i>	5	5	5
<i>Flammulina</i>	13	13	13
<i>Fomes</i>	3	3	3
<i>Fomitopsis</i>	9	9	9
<i>Galerina</i>	1	1	1
<i>Ganoderma</i>	23	22	21
<i>Gloeophyllum</i>	4	4	4
<i>Gloeoporus</i>	2	1	1
<i>Graphiola</i>	4	4	4
<i>Grifola</i>	8	8	7
<i>Gymnopilus</i>	7	7	7
<i>Hapalopilus</i>	1	1	1
<i>Hebeloma</i>	5	5	4
<i>Helicobasidium</i>	12	12	11
<i>Hericum</i>	7	7	7
<i>Heterobasidion</i>	3	3	3
<i>Hydnochaete</i>	3	3	3
<i>Hygrophoropsis</i>	1	1	1
<i>Hymenochaete</i>	2	2	2
<i>Hypsizygus</i>	1	1	1
<i>Inonotus</i>	11	11	10
<i>Irpex</i>	4	4	4
<i>Ischnoderma</i>	4	2	2
<i>Junghuhnia</i>	2	2	2
<i>Kuehneromyces</i>	2	2	2

Table 1. continued.

Fungal genus	Tested <sup>(a)</sup>	1 M <sup>(b)</sup>	1 Y <sup>(b)</sup>
<i>Laccaria</i>	3	3	2
<i>Lactarius</i>	3	3	3
<i>Laeticorticium</i>	1	1	1
<i>Laetiporus</i>	12	12	12
<i>Laetisaria</i>	1	1	1
<i>Lampteromyces</i>	10	10	10
<i>Lanopila</i>	1	1	1
<i>Lentinellus</i>	1	1	1
<i>Lentinula</i>	8	8	8
<i>Lentinus</i>	8	8	8
<i>Lenzites</i>	3	3	3
<i>Lepiota</i>	3	3	3
<i>Lepista</i>	6	5	5
<i>Leucoagaricus</i>	4	3	3
<i>Leucocoprinus</i>	3	3	3
<i>Leucopaxillus</i>	3	3	2
<i>Linderia</i>	1	1	1
<i>Loweoporus</i>	6	6	6
<i>Lycoperdon</i>	1	1	1
<i>Lyophyllum</i>	12	12	11
<i>Macrolepiota</i>	6	6	6
<i>Marasmius</i>	20	20	20
<i>Microporus</i>	1	1	1
<i>Mundkurella</i>	4	4	4
<i>Mycena</i>	9	9	9
<i>Mycoacia</i>	2	2	2
<i>Mycoleptodomoides</i>	5	5	5
<i>Naematoloma</i>	13	13	13
<i>Neolentinus</i>	7	7	7
<i>Nia</i>	10	10	10
<i>Oligoporus</i>	6	6	6
<i>Omphalotus</i>	2	2	2
<i>Oudemansiella</i>	16	16	14
<i>Oxyporus</i>	1	1	1
<i>Panaeolina</i>	2	2	2
<i>Panaeolus</i>	2	2	2
<i>Panellus</i>	10	10	10
<i>Panus</i>	3	3	3
<i>Paxillus</i>	3	3	3
<i>Pellicularia</i>	1	1	1
<i>Perenniporia</i>	10	10	10
<i>Phaeolepiota</i>	3	3	2
<i>Phaeolus</i>	5	5	5
<i>Phellinus</i>	51	51	51
<i>Phlebia</i>	4	4	4
<i>Phleogena</i>	33	33	33
<i>Pholiota</i>	18	18	17

Table 1. continued.

Fungal genus	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>
<i>Phyllotopsis</i>	1	1	1
<i>Piptoporus</i>	8	8	8
<i>Platygløea</i>	12	12	12
<i>Pleurocybella</i>	5	5	5
<i>Pleurotus</i>	43	42	42
<i>Plicaturopsis</i>	2	2	2
<i>Polyporellus</i>	5	5	5
<i>Polyporus</i>	18	18	17
<i>Poria</i>	1	1	1
<i>Protodaedalea</i>	2	2	1
<i>Psathyrella</i>	1	1	1
<i>Pseudochitocybe</i>	1	1	1
<i>Pseudocolus</i>	2	2	2
<i>Pseudomerulius</i>	1	1	1
<i>Psilocybe</i>	5	5	4
<i>Pulcherricium</i>	2	2	2
<i>Pycnoporus</i>	8	8	8
<i>Ramaria</i>	1	1	1
<i>Rhodophyllus</i>	1	1	1
<i>Rhodotus</i>	1	1	1
<i>Rigidoporus</i>	7	7	6
<i>Ripartitella</i>	1	1	1
<i>Russula</i>	1	1	1
<i>Schizophyllum</i>	6	6	6
<i>Schizopora</i>	2	2	2
<i>Sebacina</i>	15	15	15
<i>Serpula</i>	2	2	2
<i>Sparassis</i>	4	4	4
<i>Sporisorium</i>	5	5	5
<i>Steccherinum</i>	3	3	3
<i>Stereum</i>	7	7	7
<i>Strobilurus</i>	3	3	3
<i>Stropharia</i>	3	3	3
<i>Suillus</i>	5	4	4
<i>Thanatephorus</i>	1	1	1
<i>Tilletia</i>	3	3	3
<i>Tinctoporellus</i>	2	2	2
<i>Trametes</i>	22	22	22
<i>Tremella</i>	2	2	2
<i>Trichaphum</i>	6	6	6
<i>Tricholoma</i>	87	87	83
<i>Typhula</i>	28	28	28
<i>Tyromyces</i>	1	1	1
<i>Ustilago</i>	21	20	20
<i>Volvariella</i>	1	1	0
<i>Waitea</i>	20	20	20
<i>Wolfiporia</i>	3	3	3

Table 1. continued.

Fungal genus	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>
<i>Xeromphalina</i>	10	10	10
<i>Xerula</i>	2	2	2
<i>Xylobolus</i>	3	3	3
Anamorphic fungi			
<i>Acremonium</i>	5	5	5
<i>Acrodictys</i>	1	1	1
<i>Albophoma</i>	1	1	1
<i>Alternaria</i>	65	65	65
<i>Apiocarpella</i>	2	2	2
<i>Arthrimum</i>	17	17	17
<i>Arihrobotrys</i>	3	3	3
<i>Arthrographis</i>	2	2	2
<i>Ascochyta</i>	13	13	13
<i>Aspergillus</i>	78	78	78
<i>Asperisporium</i>	2	2	2
<i>Asterosporium</i>	1	1	1
<i>Aureobasidium</i>	11	11	11
<i>Bartalinia</i>	1	1	1
<i>Beauveria</i>	12	12	12
<i>Beltrania</i>	1	1	1
<i>Bipolaris</i>	94	94	94
<i>Botryodiplodia</i>	2	2	2
<i>Botryotrichum</i>	3	3	3
<i>Botrytis</i>	31	31	31
<i>Camarosporium</i>	1	1	1
<i>Camposporium</i>	1	1	1
<i>Catinula</i>	1	1	1
<i>Cephalophora</i>	2	2	2
<i>Cephalosporium</i>	4	4	4
<i>Cercospora</i>	74	74	72
<i>Cercosporella</i>	3	3	3
<i>Cercosporidium</i>	9	9	9
<i>Chalara</i>	2	2	2
<i>Chalaropsis</i>	1	1	1
<i>Chromelosporium</i>	3	3	3
<i>Chrysosporium</i>	1	1	1
<i>Cladorrhinum</i>	3	3	3
<i>Cladosporium</i>	61	61	61
<i>Codinaea</i>	1	1	1
<i>Colletotrichum</i>	172	172	172
<i>Coniella</i>	8	8	8
<i>Coniothyrium</i>	1	1	1
<i>Corynespora</i>	43	43	43
<i>Coryneum</i>	1	1	1
<i>Cristulariella</i>	2	2	2
<i>Cryptosporiopsis</i>	10	10	10
<i>Curvularia</i>	42	42	42

**Table 1. continued.**

Fungal genus	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>
<i>Cylindrocarpon</i>	17	17	17
<i>Cylindrocladiella</i>	3	3	3
<i>Cylindrocladium</i>	30	30	30
<i>Cylindrosporium</i>	1	1	1
<i>Cytophoma</i>	1	1	1
<i>Cytospora</i>	18	18	18
<i>Dactylaria</i>	1	1	1
<i>Dematophora</i>	4	4	4
<i>Dendrodochium</i>	1	1	1
<i>Dendryphion</i>	1	1	1
<i>Dinemasporium</i>	1	1	1
<i>Diplodia</i>	7	7	7
<i>Diplodina</i>	2	2	2
<i>Discosia</i>	4	4	4
<i>Discula</i>	1	1	1
<i>Doratomyces</i>	1	1	1
<i>Dothiorella</i>	8	8	8
<i>Dothistroma</i>	2	2	2
<i>Drechslera</i>	50	50	50
<i>Duosporium</i>	2	2	2
<i>Embellisia</i>	1	1	1
<i>Endothiella</i>	1	1	1
<i>Epicoccum</i>	17	17	17
<i>Exserohilum</i>	15	15	15
<i>Fulvia</i>	11	11	11
<i>Fusarium</i>	633	633	633
<i>Geosmithia</i>	1	1	1
<i>Gliocladium</i>	10	10	10
<i>Gloeocercospora</i>	11	11	11
<i>Haprosoporella</i>	1	1	1
<i>Harzia</i>	5	5	5
<i>Helicomycetes</i>	1	1	1
<i>Helminthosporium</i>	4	4	4
<i>Hendersonula</i>	1	1	1
<i>Humicola</i>	11	11	11
<i>Hyalodendron</i>	1	1	1
<i>Hymenula</i>	1	1	1
<i>Hyphodiscosia</i>	1	1	1
<i>Irpicomycetes</i>	1	1	1
<i>Kabatiella</i>	2	2	2
<i>Lasiodiplodia</i>	16	16	16
<i>Leptochlamys</i>	1	1	1
<i>Leptographium</i>	2	2	2
<i>Macrophoma</i>	20	20	20
<i>Macrophomina</i>	17	17	17
<i>Mammaria</i>	1	1	1
<i>Mariannaea</i>	3	3	3

**Table 1. continued.**

Fungal genus	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>
<i>Marssonina</i>	5	5	5
<i>Melanconium</i>	7	7	7
<i>Metarhizium</i>	9	9	9
<i>Microdochium</i>	30	30	30
<i>Monacrosporium</i>	4	4	4
<i>Monilia</i>	4	4	4
<i>Monochaetia</i>	2	2	2
<i>Monocillium</i>	1	1	1
<i>Monostichella</i>	1	1	1
<i>Monotosporella</i>	1	1	1
<i>Myceliophthora</i>	1	1	1
<i>Mycocentrospora</i>	2	2	2
<i>Mycovellosiella</i>	2	2	2
<i>Myrothecium</i>	11	11	11
<i>Myxosporium</i>	1	1	1
<i>Naranus</i>	1	1	1
<i>Nigroporus</i>	1	1	1
<i>Nigrospora</i>	31	31	31
<i>Nodulisporium</i>	3	3	3
<i>Nomuraea</i>	3	3	3
<i>Ochroconis</i>	5	5	5
<i>Oedocephalum</i>	1	1	1
<i>Oidiodendron</i>	10	10	10
<i>Paecilomyces</i>	11	11	11
<i>Papulaspora</i>	3	3	3
<i>Paracercospora</i>	2	2	2
<i>Penicillium</i>	9	9	9
<i>Periconia</i>	5	5	5
<i>Pestalotia</i>	17	17	17
<i>Pestalotiopsis</i>	107	107	107
<i>Phaeoisariopsis</i>	3	3	3
<i>Phaeoseptoria</i>	4	4	4
<i>Phialomyces</i>	1	1	1
<i>Phialophora</i>	17	17	17
<i>Phloeospora</i>	2	2	2
<i>Phoma</i>	40	40	40
<i>Phomopsis</i>	84	84	84
<i>Phyllosticta</i>	30	30	30
<i>Pithomyces</i>	6	6	6
<i>Pleiochaeta</i>	2	2	2
<i>Pseudocercospora</i>	74	74	74
<i>Pseudocercosporella</i>	37	37	37
<i>Pseudoseptoria</i>	1	1	0
<i>Pyrenochaeta</i>	30	30	30
<i>Pyricularia</i>	111	111	111
<i>Racodium</i>	1	1	1
<i>Raffaelea</i>	3	3	3

Table 1. continued.

Fungal genus	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>
<i>Ramichloridium</i>	12	12	12
<i>Ramularia</i>	2	2	2
<i>Rhizoctonia</i>	241	234	227
<i>Rhizosphaera</i>	9	9	9
<i>Rhynchosporium</i>	7	7	7
<i>Robillarda</i>	2	2	2
<i>Sarocladium</i>	1	1	1
<i>Sclerotium</i>	43	43	42
<i>Scopulariopsis</i>	2	2	2
<i>Seimatosporium</i>	2	2	2
<i>Seiridium</i>	12	12	12
<i>Selenophoma</i>	3	3	3
<i>Septogloeum</i>	1	1	1
<i>Septonema</i>	2	2	2
<i>Septoria</i>	16	16	16
<i>Septotis</i>	3	3	3
<i>Sirosporium</i>	2	2	2
<i>Spegazzinia</i>	1	1	1
<i>Sphaceloma</i>	6	6	5
<i>Sphaerellopsis</i>	48	48	48
<i>Sphaeropsis</i>	8	8	8
<i>Sporidesmium</i>	6	6	6
<i>Stachybotrys</i>	2	2	2
<i>Stagonospora</i>	4	4	4
<i>Staphylotrichum</i>	3	3	3
<i>Stemphylium</i>	10	10	10
<i>Stigmina</i>	6	6	6
<i>Strasserioopsis</i>	2	2	2

Table 1. continued.

Fungal genus	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>
<i>Taeniolella</i>	1	1	1
<i>Tetracladium</i>	2	2	2
<i>Thielaviopsis</i>	3	3	3
<i>Thysanophora</i>	1	1	1
<i>Ticogloea</i>	1	1	1
<i>Torula</i>	4	4	4
<i>Torulomyces</i>	1	1	1
<i>Trichocladium</i>	1	1	1
<i>Trichoderma</i>	28	28	28
<i>Trichothecium</i>	4	4	4
<i>Trinacrium</i>	1	1	1
<i>Triposphermum</i>	1	1	1
<i>Tritirachium</i>	1	1	1
<i>Trochophora</i>	7	7	7
<i>Truncatella</i>	1	1	1
<i>Tubakia</i>	4	4	4
<i>Tubercularia</i>	3	3	3
<i>Ulocladium</i>	6	6	6
<i>Ustilaginoidea</i>	2	1	1
<i>Verticillium</i>	152	152	152
<i>Volutella</i>	2	2	2
<i>Wallemia</i>	1	1	1
Total	5,651	5,503	5,406

a) Number of strains tested.

b) Number of strains surviving for a period indicated, 1 month (1M) or 1 year (1Y).

Table 2. Fungal species not showing 100% recovery

Fungal species	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>	Recovery <sup>c)</sup>
Oomycota				
<i>Achlya americana</i>	2	0	0	0
<i>Achlya caroliniana</i>	1	0	0	0
<i>Achlya conspicua</i>	2	0	0	0
<i>Achlya flagellata</i>	6	1	0	0
<i>Achlya klebsiana</i>	1	0	0	0
<i>Achlya</i> sp.	2	0	0	0
<i>Achlya stellata</i>	1	0	0	0
<i>Aphanomyces cladogamus</i>	1	0	0	0
<i>Aphanomyces cochlioides</i>	3	1	1	33
<i>Aphanomyces euteiches</i>	1	0	0	0
<i>Dictyuchus anomalus</i>	2	0	0	0
<i>Dictyuchus carphophorus</i>	1	0	0	0
<i>Dictyuchus pseudoachlyoides</i>	1	0	0	0
<i>Dictyuchus sterile</i>	2	0	0	0

**Table 2. continued.**

Fungal species	Tested <sup>(a)</sup>	1 M <sup>(b)</sup>	1 Y <sup>(b)</sup>	Recovery <sup>(c)</sup>
<i>Phytophthora cactorum</i>	7	6	6	86
<i>Phytophthora capsici</i>	5	2	2	40
<i>Phytophthora infestans</i>	15	11	11	73
<i>Phytophthora melonis</i>	1	0	0	0
<i>Phytophthora nicotianae</i>	35	32	27	77
<i>Phytophthora porri</i>	2	1	0	0
<i>Phytophthora</i> sp.	15	13	10	67
<i>Phytophthora vignae</i>	6	5	4	67
<i>Pythium aphanidermatum</i>	21	14	11	52
<i>Pythium apteroticum</i>	1	0	0	0
<i>Pythium carolinianum</i>	6	3	2	33
<i>Pythium dissimile</i>	1	0	0	0
<i>Pythium dissotocum</i>	7	5	5	71
<i>Pythium echinulatum</i>	5	4	2	40
<i>Pythium elongatum</i>	4	3	2	50
<i>Pythium graminicola</i>	12	10	7	58
<i>Pythium hydno sporum</i>	4	1	1	25
<i>Pythium inflatum</i>	6	4	4	67
<i>Pythium intermedium</i>	9	8	8	89
<i>Pythium irregulare</i>	28	26	24	86
<i>Pythium myriotylum</i>	1	0	0	0
<i>Pythium oedo chylum</i>	7	2	0	0
<i>Pythium periplocum</i>	2	1	1	50
<i>Pythium rostratum</i>	9	5	5	56
<i>Pythium</i> sp.	7	4	4	57
<i>Pythium spinosum</i>	23	15	11	48
<i>Pythium splendens</i>	9	3	0	0
<i>Pythium sylvaticum</i>	10	10	8	80
<i>Pythium torulosum</i>	25	18	16	64
<i>Pythium ultimum</i>	27	14	11	41
<i>Pythium vexans</i>	4	0	0	0
<i>Thraustotheca clavata</i>	4	1	1	25
Zygomycota				
<i>Choanephora infundibulifera</i>	2	1	1	50
Ascomycota				
<i>Cistella japonica</i>	19	19	16	84
<i>Gremmeniella abietina</i>	9	9	8	89
<i>Halosphaeria appendiculata</i>	18	18	17	94
<i>Morchella esculenta</i>	3	3	2	67
<i>Peziza ostracoderma</i>	2	2	0	0
<i>Peziza vesiculosa</i>	2	1	1	50
<i>Remispora stellata</i>	16	16	15	94
<i>Sclerotinia sclerotiorum</i>	29	29	28	97
<i>Tricholoma bakamatsutake</i>	10	10	9	90
<i>Tricholoma matsutake</i>	59	59	56	95
Basidiomycota				
<i>Amanita muscaria</i>	1	1	0	0



Table 2. continued.

Fungal species	Tested <sup>a)</sup>	1 M <sup>b)</sup>	1 Y <sup>b)</sup>	Recovery <sup>c)</sup>
<i>Armillaria mellea</i> ( <i>sensu lato</i> )	41	40	40	98
<i>Armillaria</i> sp.	2	1	1	50
<i>Auricularia auricula</i>	4	4	3	75
<i>Auricularia polytricha</i>	11	9	8	73
<i>Ceratobasidium cornigerum</i>	8	6	6	75
<i>Ceratobasidium setariae</i>	9	9	8	89
<i>Ceratobasidium</i> sp.	22	21	13	59
<i>Coprinus atramentarius</i>	1	1	0	0
<i>Eichleriella leucophaea</i>	10	10	8	80
<i>Erythricium salmonicolor</i>	18	17	17	94
<i>Ganoderma applanatum</i> ( <i>sensu lato</i> )	9	8	8	89
<i>Ganoderma lucidum</i>	6	6	5	83
<i>Gloeoporus dichrous</i>	2	1	1	50
<i>Grifola frondosa</i>	8	8	7	88
<i>Hebeloma spoliatum</i>	2	2	1	50
<i>Helicobasidium mompa</i>	12	12	11	92
<i>Inonotus obliquus</i>	4	4	3	75
<i>Ischnoderma resinosum</i>	4	2	2	50
<i>Laccaria laccata</i>	1	1	0	0
<i>Lepista nuda</i>	3	2	2	67
<i>Leucoagaricus rubrotinctus</i>	1	0	0	0
<i>Leucopaxillus septentrionalis</i>	2	2	1	50
<i>Lyophyllum sykosporum</i>	1	1	0	0
<i>Oudemansiella pudens</i>	2	2	1	50
<i>Oudemansiella radicata</i>	7	7	6	86
<i>Phaeolepiota aurea</i>	3	3	2	67
<i>Pholiota lucifera</i>	1	1	0	0
<i>Pleurotus eugrammus</i>	1	0	0	0
<i>Polyporus badius</i>	1	1	0	0
<i>Protodaedalea hispida</i>	2	2	1	50
<i>Psilocybe argentipes</i>	3	3	2	67
<i>Rigidoporus ulmarius</i>	2	2	1	50
<i>Ustilago esculenta</i>	7	6	6	86
<i>Volvariella volvacea</i>	1	1	0	0
Anamorphic fungi				
<i>Cercospora citrullina</i>	2	2	1	50
<i>Cercospora corchori</i>	1	1	0	0
<i>Pseudoseptoria donacis</i>	1	1	0	0
<i>Rhizoctonia solani</i>	160	156	150	94
<i>Rhizoctonia</i> sp. (binucleate)	64	63	62	97
<i>Rhizoctonia</i> sp.	6	4	4	67
<i>Sclerotium rolfsii</i>	29	29	28	97
<i>Sphaceloma</i> sp.	3	3	2	67
<i>Ustilaginoidea virens</i>	2	1	1	50

a) Number of tested strains.

b) Number of strains surviving for a period indicated, 1 month (1M) or 1 year (1Y).

c) Percentage of strains surviving during the 1-year storage.

“Fluctuation” has been reported in the preservation of some strains: i.e., some fungal strains were not revived after storage for a certain period, but could be occasionally revived later after longer storage (2). However, information about the phenomenon was lost on data processing, because Tables 1 and 2 were made based on the longest periods of successful storage for each strain (4). An example of “fluctuation” in our study was *Phytophthora infestans* MAFF 236317, which had 10% viability after 1 month storage, but which showed 80% viability after 1 year. The cause of the “fluctuation” remains to be elucidated. One cause may be experimental error, because repeated examinations often show normal viabilities. Thus, replication of stocks is important to avoid errors or reporting “fluctuation.”

Cryopreservation after uncontrolled freezing was shown to be effective and useful for the storage of many fungal strains. This supports the use of the cryopreservation in the MAFF Genebank. Longevity of the strains preserved by the cryopreservation in the Genebank will be surveyed further.

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### 未制御で凍結した糸状菌類の液体窒素気相中での保存

永井利郎, 井手野亜伊香, 柘植倫代, 小柳千栄, 鬼木正臣, 喜多孝一,  
堀田光生, 青木孝之, 小林享夫, 土屋健一

農林水産省農業生物資源研究所

農林水産省ジーンバンクでは, 糸状菌類 (卵菌類を含む) を冷却速度を制御しない方法で凍結した後,  $-165^{\circ}\text{C}$  の液体窒素気相中に保存してきた。保存されている 466 属 1,182 種にわたる 5,651 菌株の生残率を調べた結果, 1 年間安定に保存できた菌株数は 5,406 株 (95.7%) であった。1 年間の保存が可能であった菌株の割合は, 卵菌門に属する菌株では 59.3% (233/393 供試菌株数), 接合菌門 98.3% (58/59), 子囊菌門 98.8% (881/892), 担子菌門 95.4% (1,100/1,153), 不完全菌類 99.4% (3,134/3,154) となった。 *Achlya*, *Dictyuchus* (卵菌門), *Volvariella* (担子菌門) および *Pseudoseptoria* (不完全菌類) 属の糸状菌類には本法は適用できなかった。